

# Techniques in Neurophysiology

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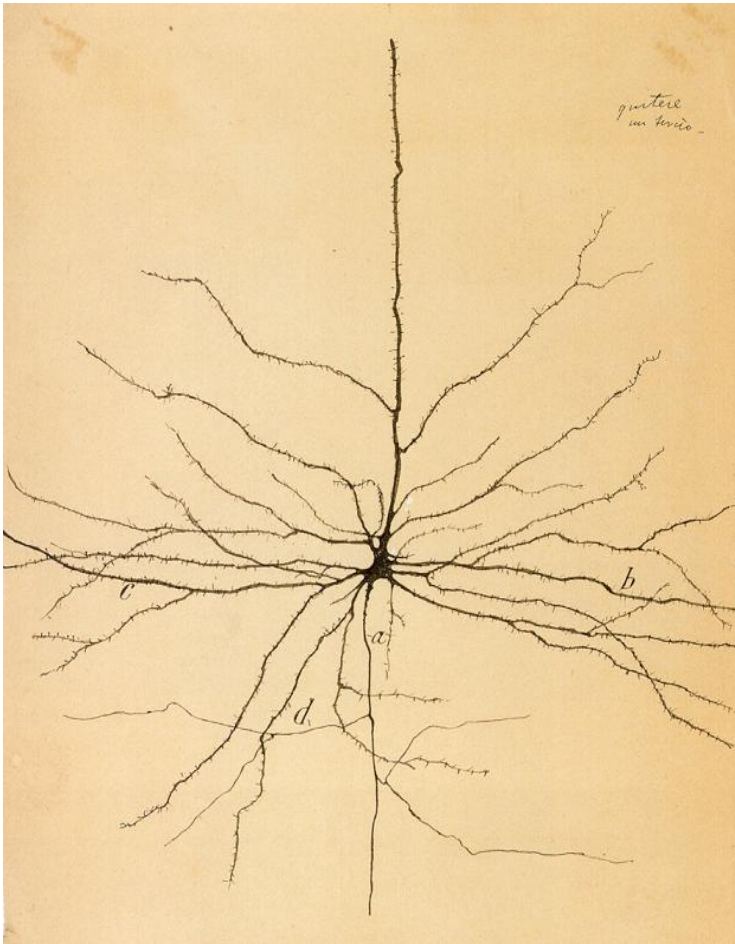
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$10^{11}$  neurons  
 $10^{12}$  glial cells  
>  $10^{15}$  connections

<http://science.nationalgeographic.com>

# Basics of neuronal function

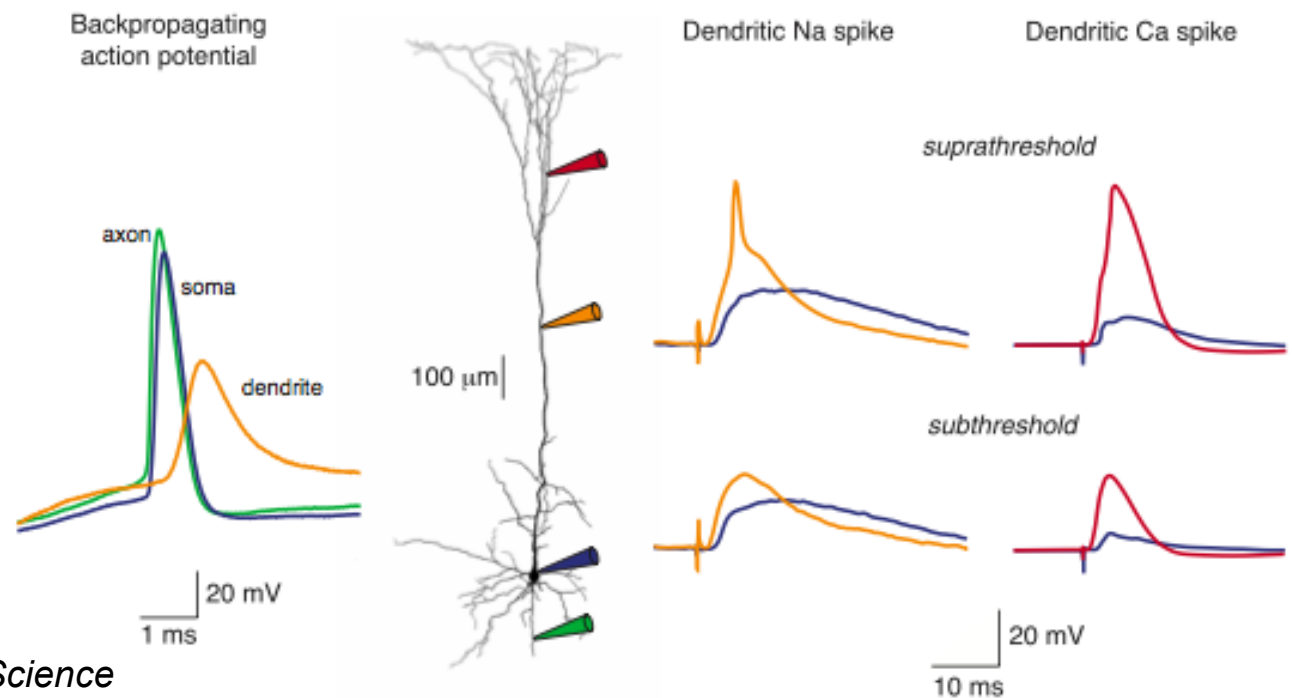


Cajal, 1899

- Cell membrane is lipid-based
- Electrochemical gradients of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$ , etc., are maintained at great expense
- Ions pass through discrete “channels” to alter membrane potential  $V_{\text{in}} - V_{\text{out}}$
- “Dendrites” collect as many as  $10^4$  inputs
- Neuronal output (soma and axon): digital pulse of amplitude 100 mV, duration 1-5 ms
- To “learn” is to:
  - Alter synaptic strength
  - Alter postsynaptic integration
- Daemons tune the system 24/7
- Dead neurons don't regenerate well
- The brain doesn't like human-made materials
- Our measurement tools are primitive
- Non-stationaries and noise are ubiquitous

# State of the art(?)

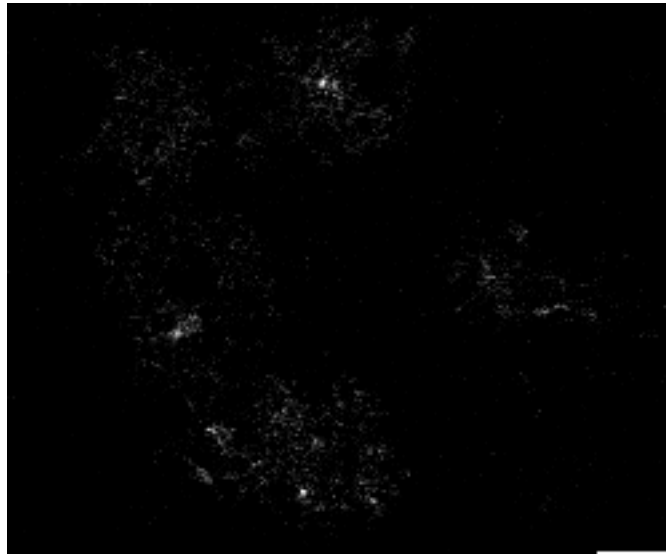
- Measuring membrane potential in one or two spots with good temporal bandwidth
- *Spatial distribution of channel densities is usually unknowable*



Häusser et al. (2000) *Science*

# State of the art(?)

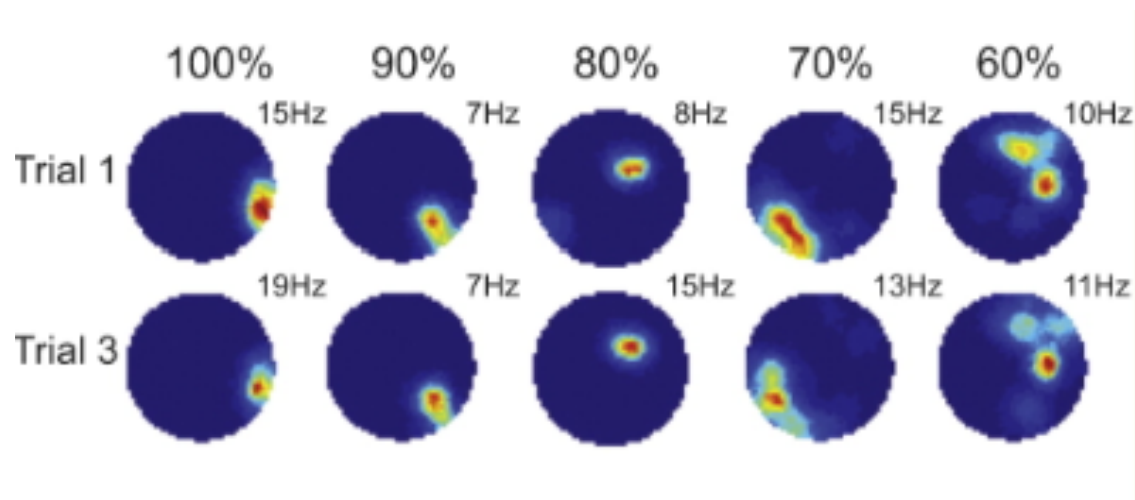
- Measuring qualitative changes in  $\text{Ca}^{2+}$  with good spatial resolution, but poor and distorted temporal resolution
- *We only have the foggiest idea about intracellular messages; the measurements we have are questionable*



Data collected by Roy Smeal, in collaboration with Mario Capecchi, Petr Tvrdek, and Karen Willcox

# State of the art(?)

- Measuring spike trains in 1–100 cells simultaneously, and making educated guesses of the cell types
- *Very difficult to parse cell types; measurements biased toward large cells located in dense populations*

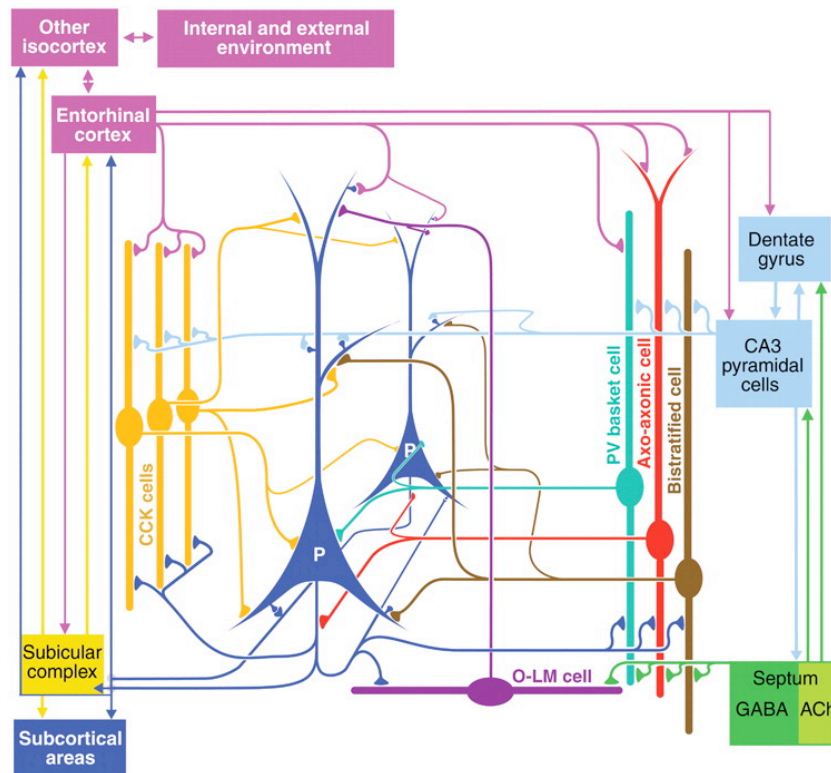


Henriksen et al. (2010) *Neuron*

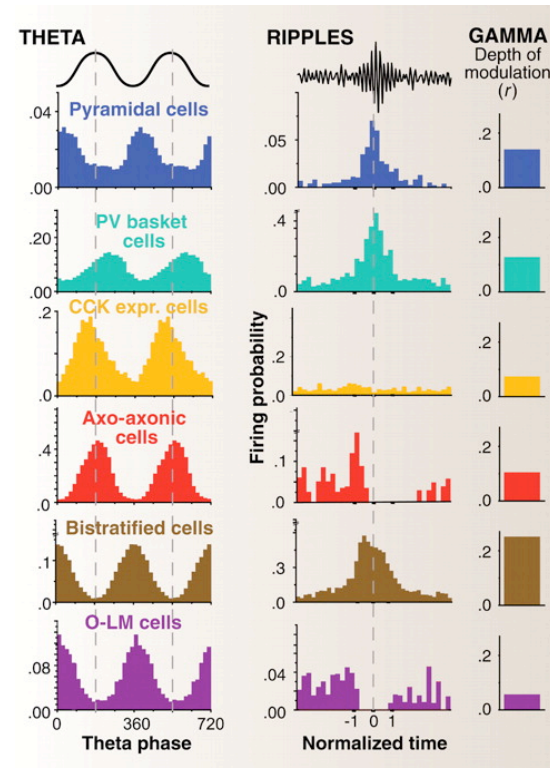


# State of the art(?)

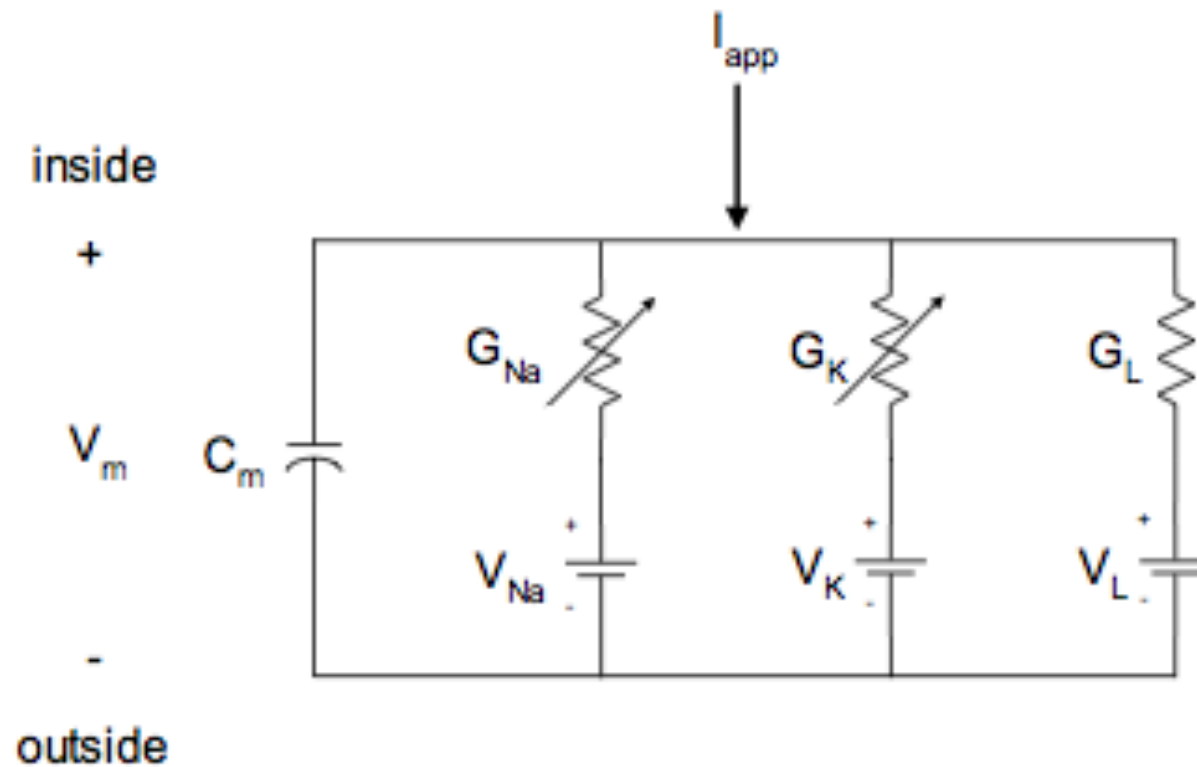
- Intracellular or juxtacellular recording and staining of cells in anesthetized animals
- *Heroic; yields low; anesthesia usually required*



Klausberger and Somogyi (2008) *Science*



# The Hodgkin-Huxley model





# The Hodgkin-Huxley model

$$C_m \frac{dV_m}{dt} = -[G_{\text{Na}}(V_m - V_{\text{Na}})$$

$$+ G_{\text{K}}(V_m - V_{\text{K}}) + G_{\text{L}}(V_m - V_{\text{L}})] + I_{\text{app}}$$

$$G_{\text{Na}} = \bar{G}_{\text{Na}} m^3 h \quad G_{\text{K}} = \bar{G}_{\text{K}} n^4$$

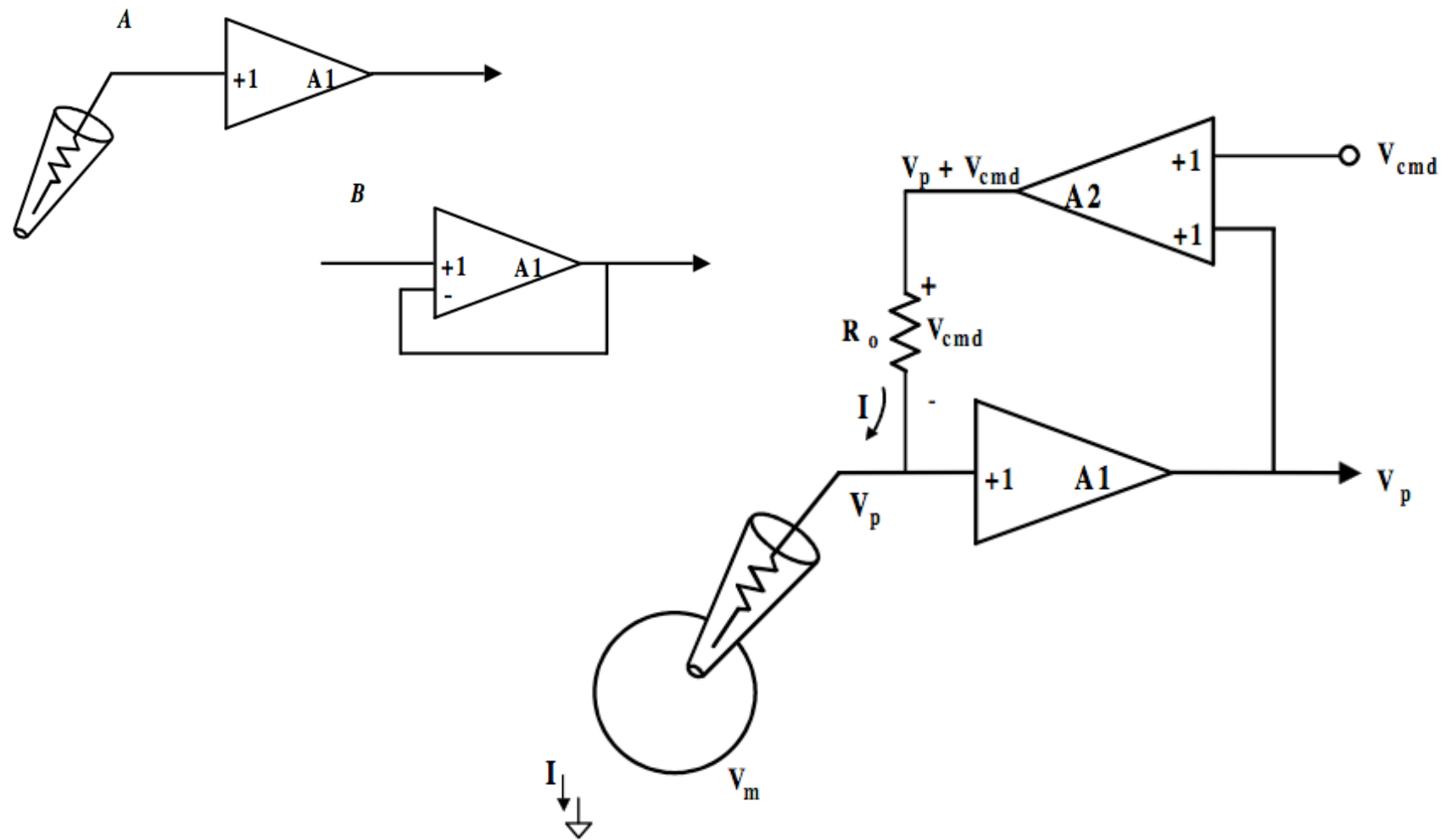
$$\frac{dm}{dt} = \frac{m_{\infty}(V_m) - m}{\tau_m(V_m)} \quad \frac{dh}{dt} = \frac{h_{\infty}(V_m) - h}{\tau_h(V_m)}$$

$$\frac{dn}{dt} = \frac{n_{\infty}(V_m) - n}{\tau_n(V_m)}$$

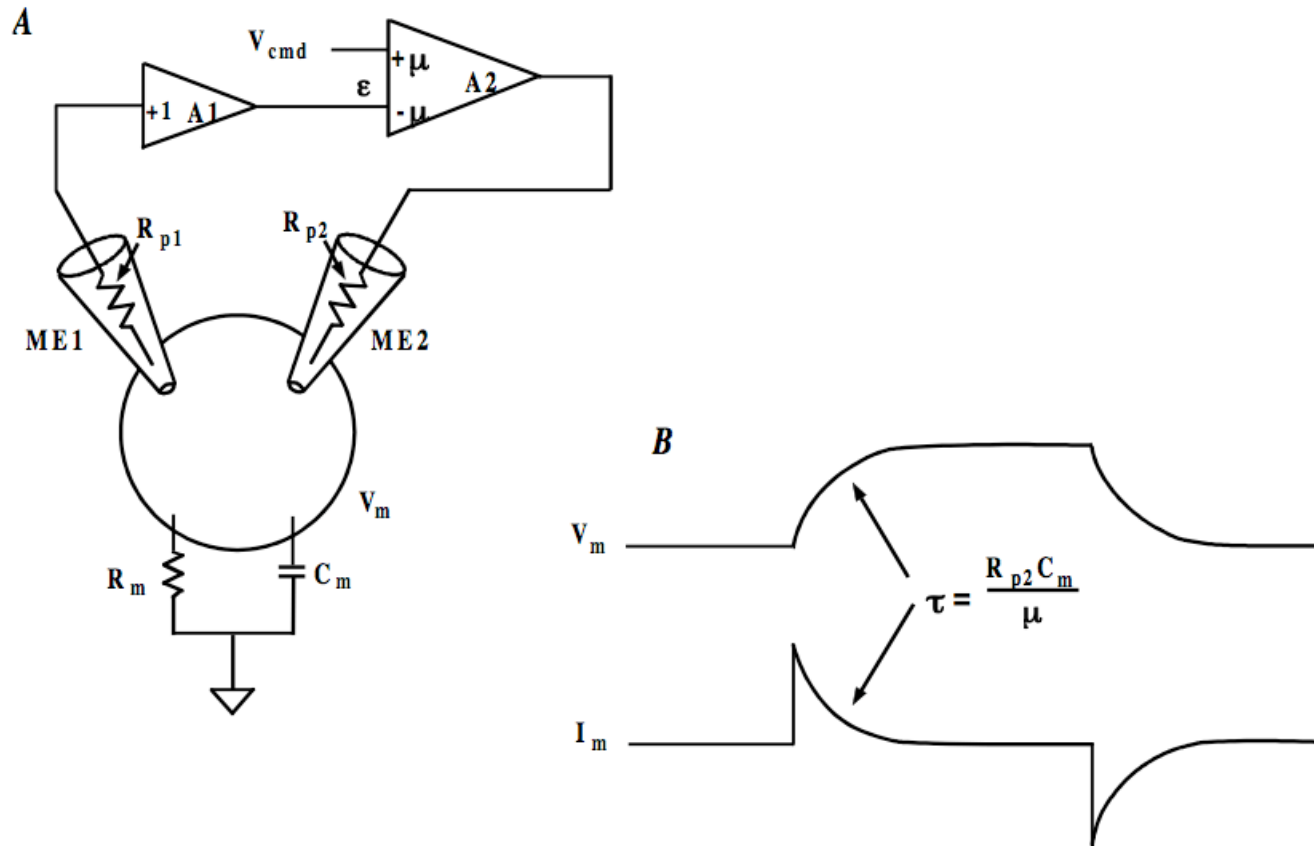
# Intracellular measurement techniques

- Current clamp – control current “injected” into cell, measure  $V_m$
- Voltage clamp – control  $V_m$ , measure amount of current required to do so
- “Dynamic” clamp – measure  $V_m$ , inject current that can represent voltage-gated channels or postsynaptic effects of virtual neurons

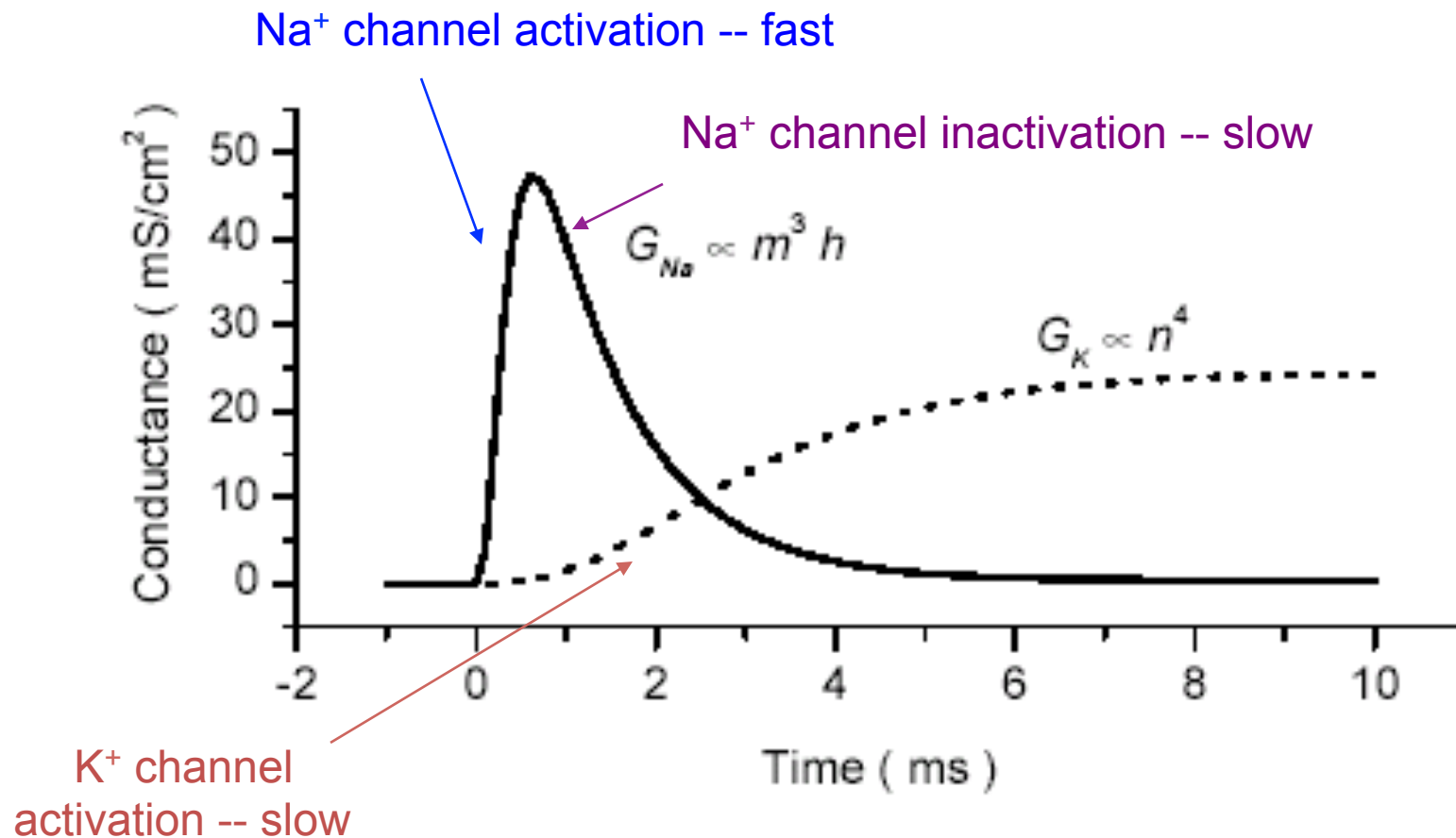
# Current clamp



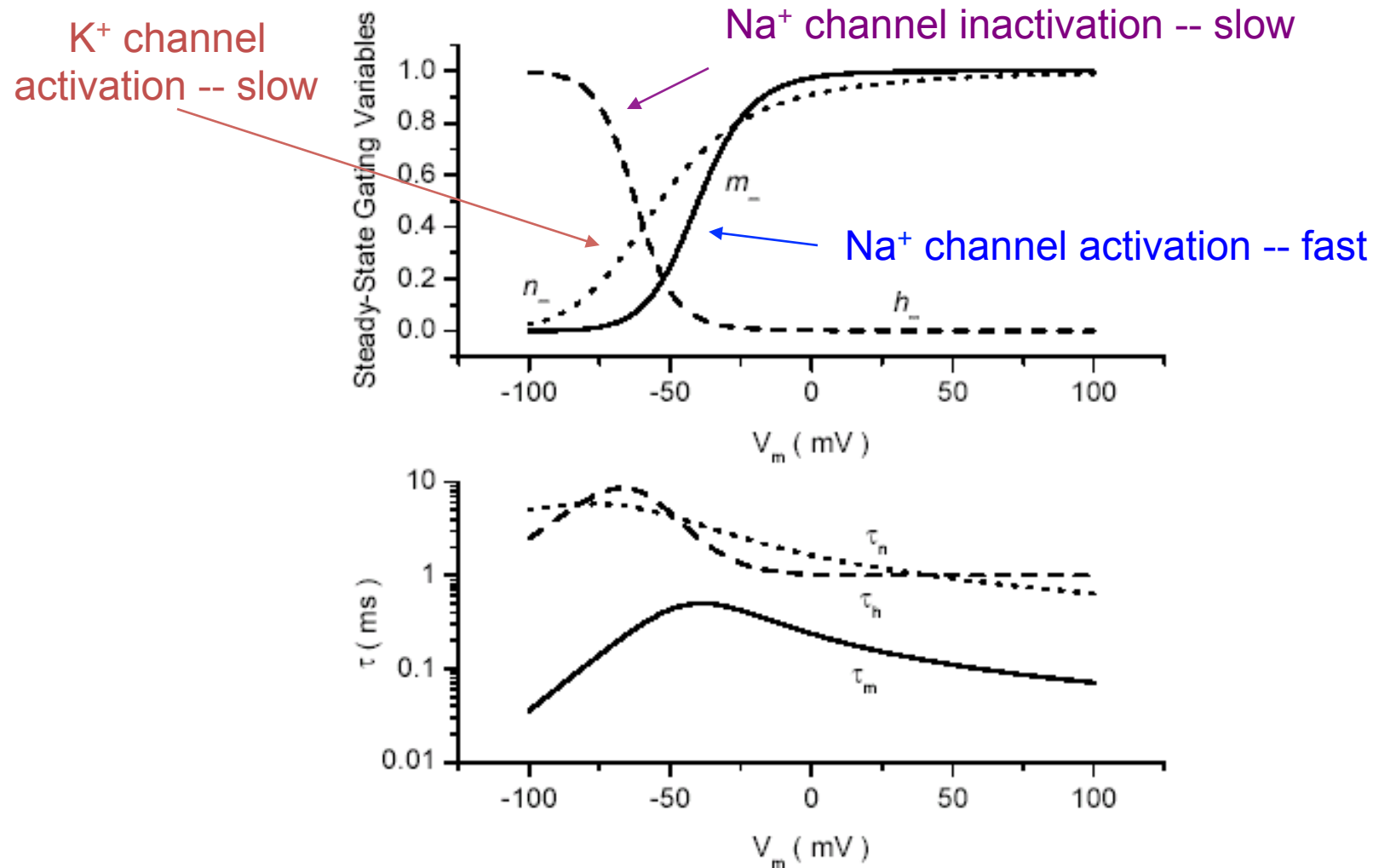
# Voltage clamp



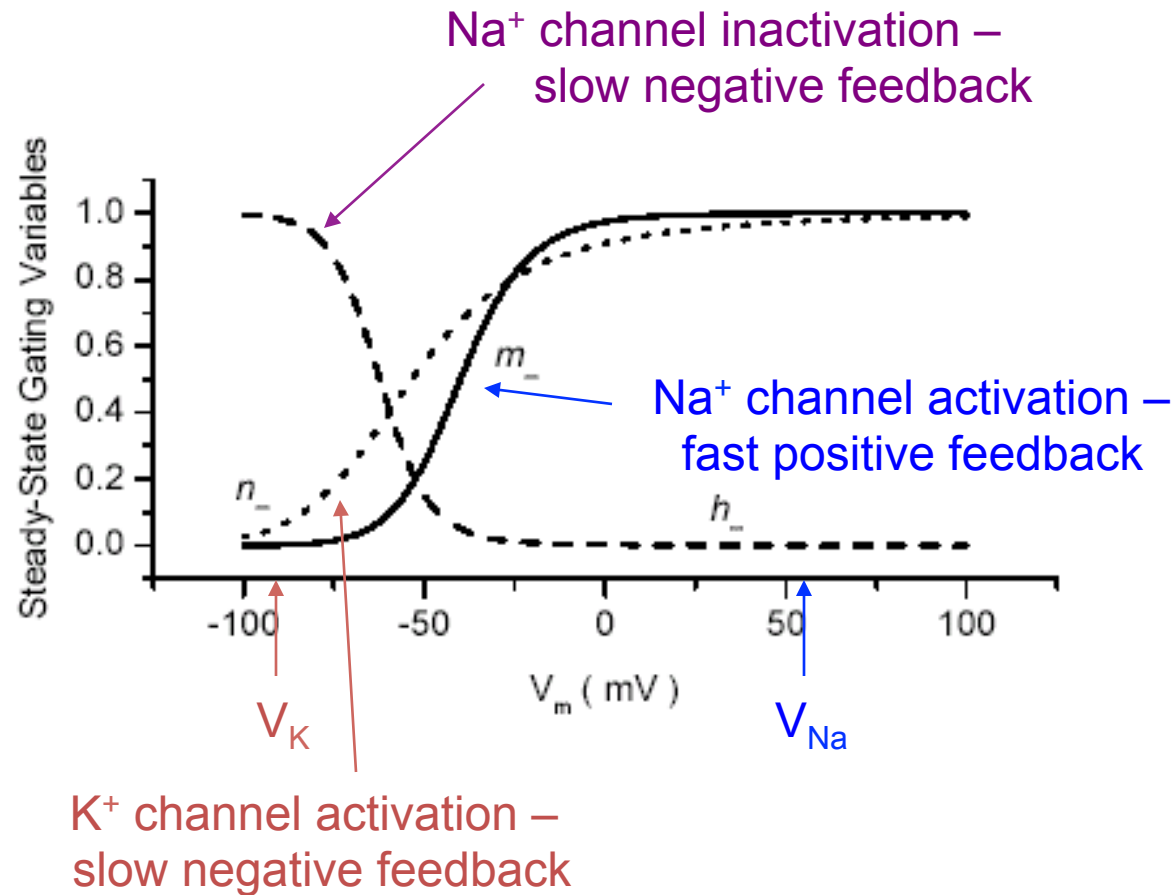
# Typical Voltage-Clamp Responses



# Parameters for Squid $\text{Na}^+$ and $\text{K}^+$ Channels

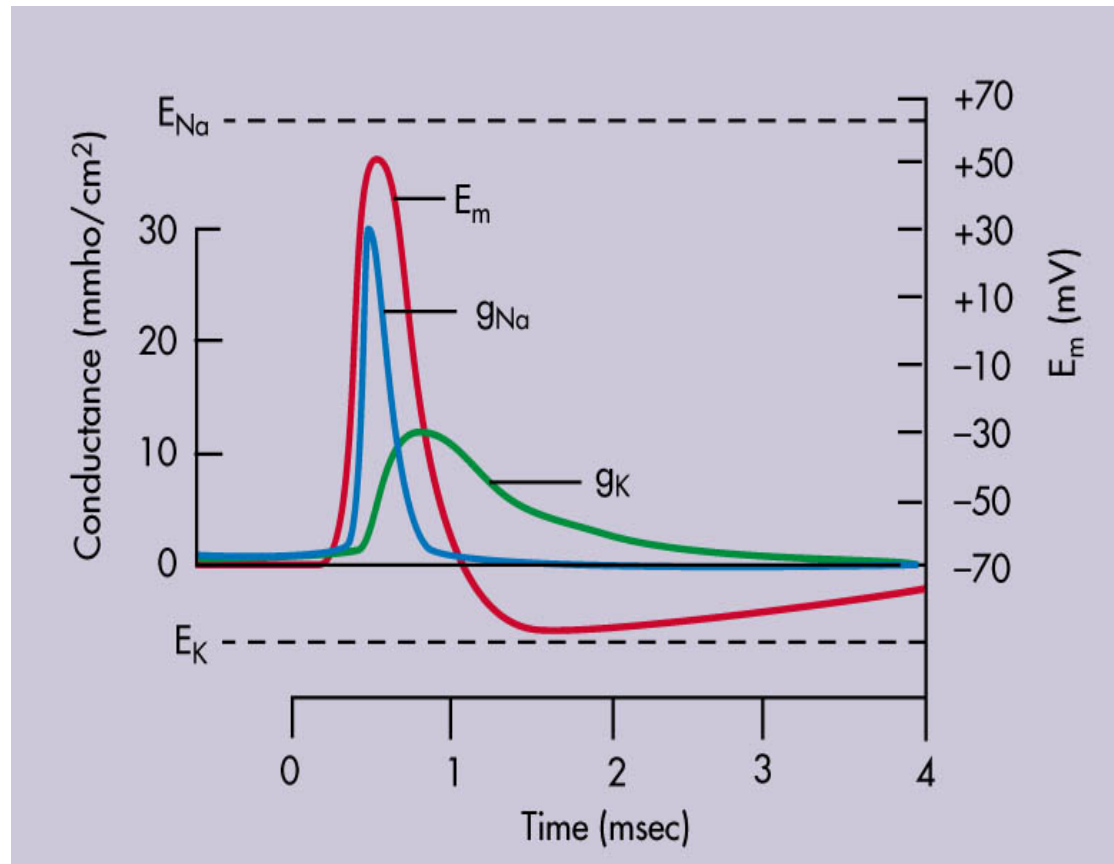


# Parameters for Squid $\text{Na}^+$ and $\text{K}^+$ Channels



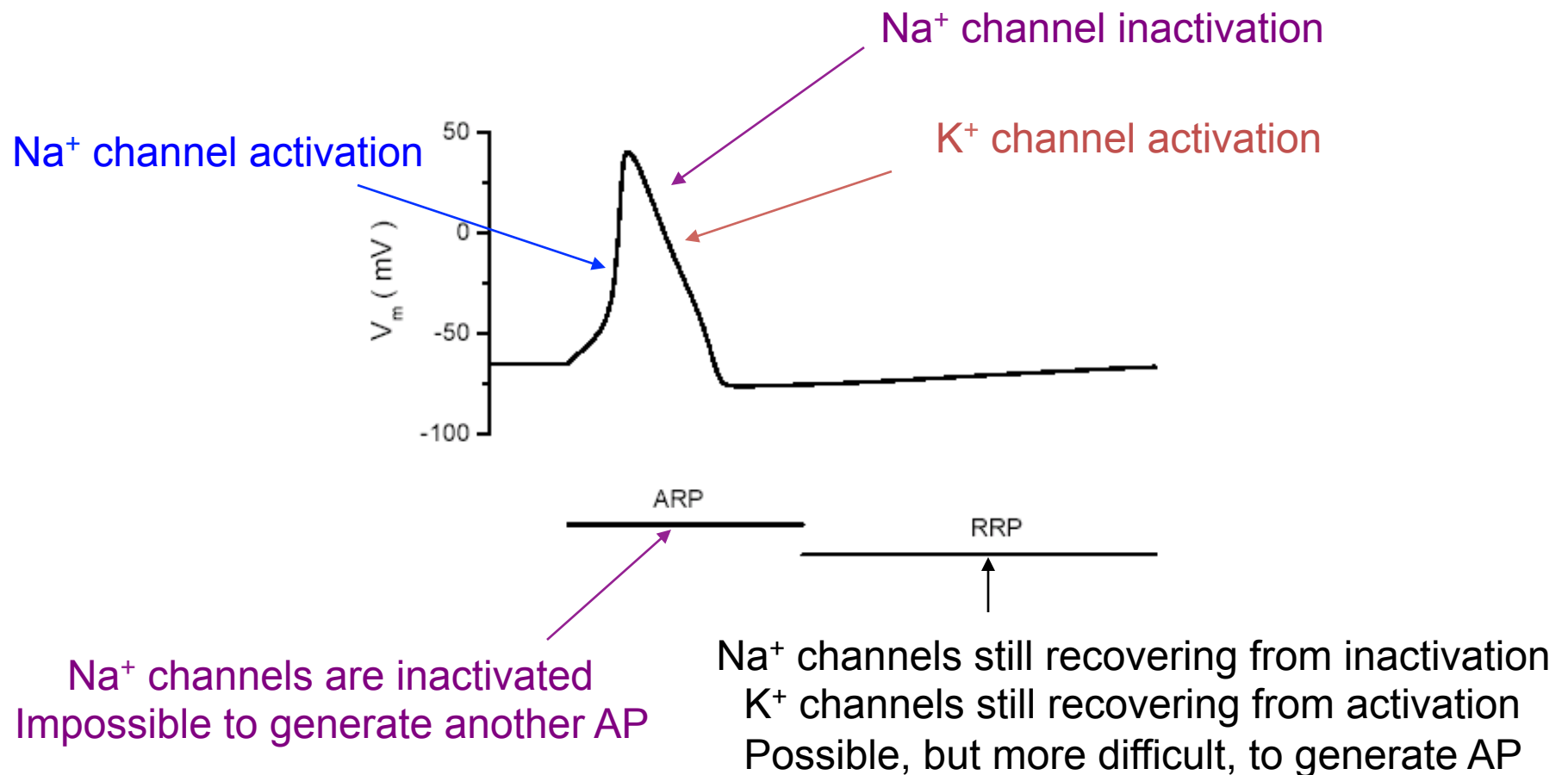


# Conductance Changes Underlying the Action Potential

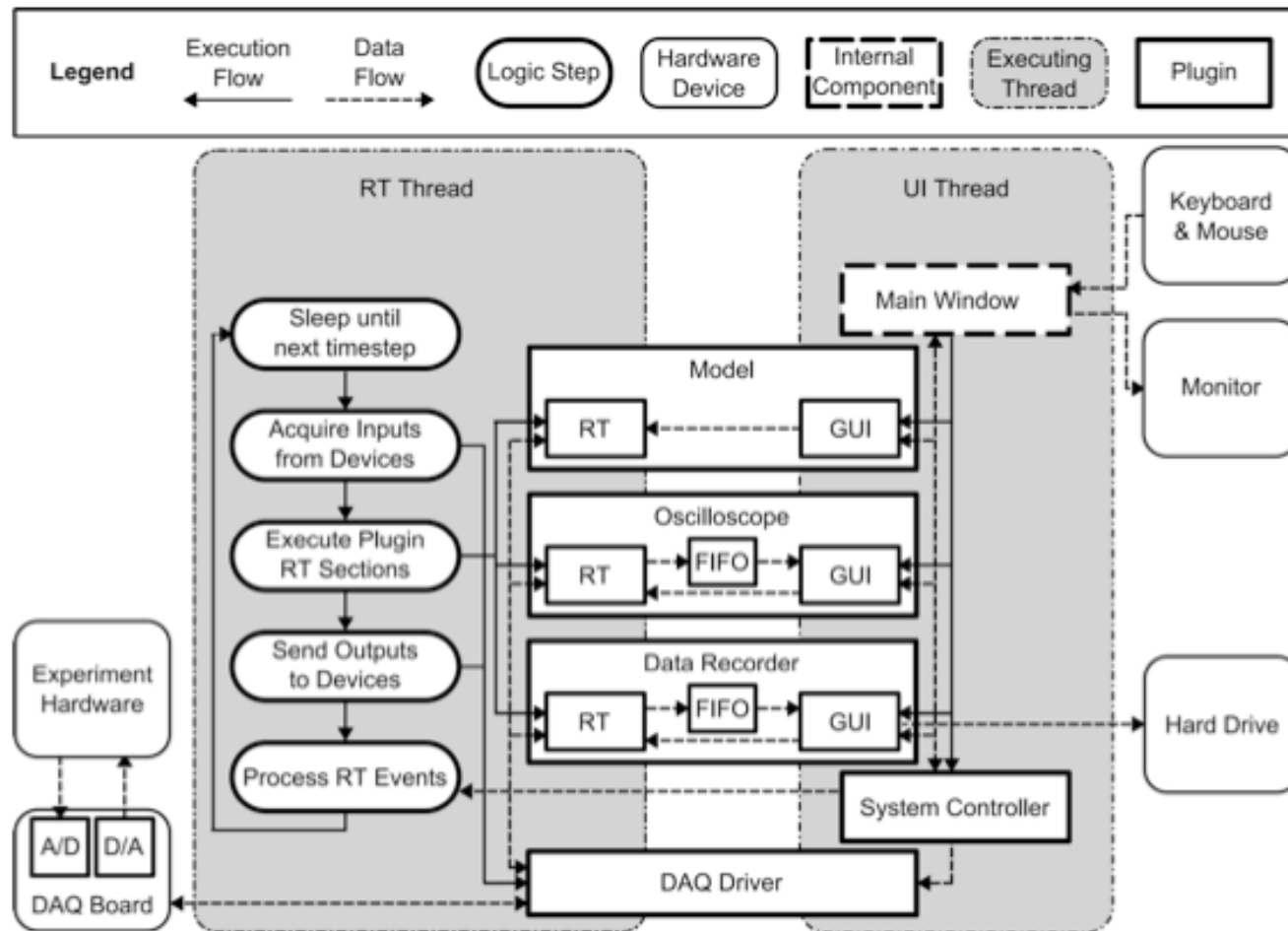


Berne and Levy

# The Action Potential Explained



# Dynamic clamp: adding arbitrary conductances to living neurons



## RTXI: Current applications

- Measuring responses to artificial synaptic inputs to build nonlinear-dynamics-inspired descriptions of cellular I-O properties and network coherence

Netoff et al. *J Neurophysiol* 93: 1197-1208, 2005; Netoff et al. *J Comput Neurosci* 18: 287-295, 2005; Pervouchine et al. *Neural Comput* 18:2617-2650, 2006; Fernandez and White *J Neurosci* 28: 3790-3803, 2008; Fernandez and White *J Neurosci* 29: 973-986, 2009; Economo et al. *J Neurosci* 30:2407-2413; Fernandez and White *J Neurosci* 30: 230-241, 2010; Fernandez et al. *J Neurosci* 31: 3880-3993, 2011.

- Altering the properties of voltage-gated channels *in vitro*

Dorval and White *J Neurosci* 25: 10025-10028, 2005; Fernandez and White *J Neurosci* 28: 3790-3803, 2008; Fernandez and White *J Neurosci* 29: 973-986, 2009; Economo et al. *J Neurosci* 30:2407-2413; Fernandez and White *J Neurosci* 30: 230-241, 2010; Fernandez et al. *J Neurosci* 31: 3880-3993, 2011

- Virtual acoustic environments for studying the use of dynamic cues in sound localization

Scarpaci et al. *Proc ICAD* 05, 214-246

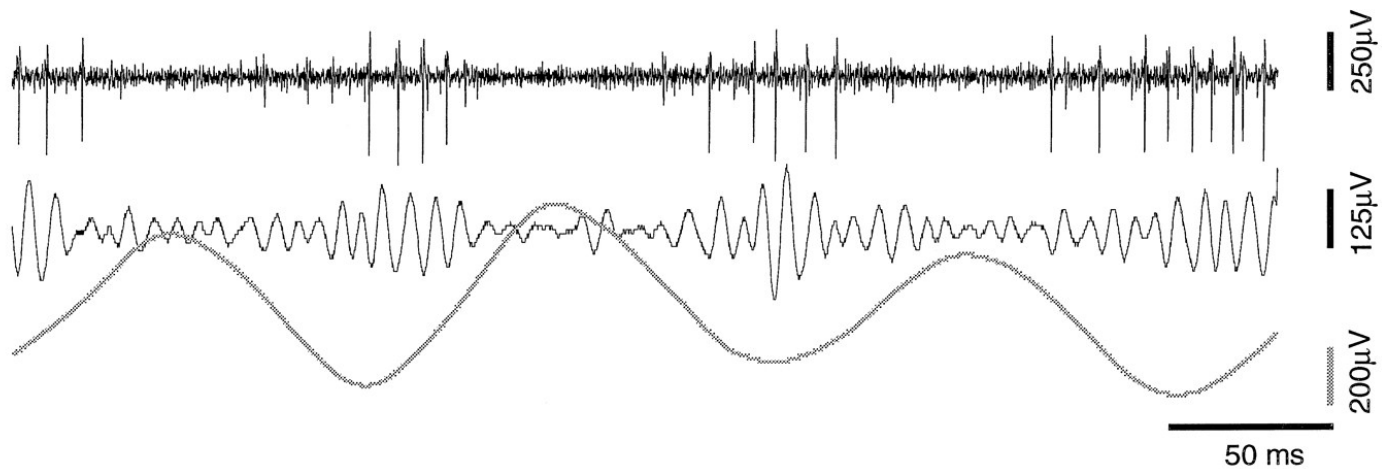
# What does it mean to “inject” conductance?

$$I_{virtual}(t) = G_{virtual}(t) (V_m(t) - E_{rev,virtual})$$

Typical choices of  $G_{virtual}$ :

- Poisson-process-driven synaptic waveforms
- Solutions from HH-style gating equations
- Synaptic waveforms driven by spikes in a biological cell

# Synchronous rhythmic activity in hippocampus



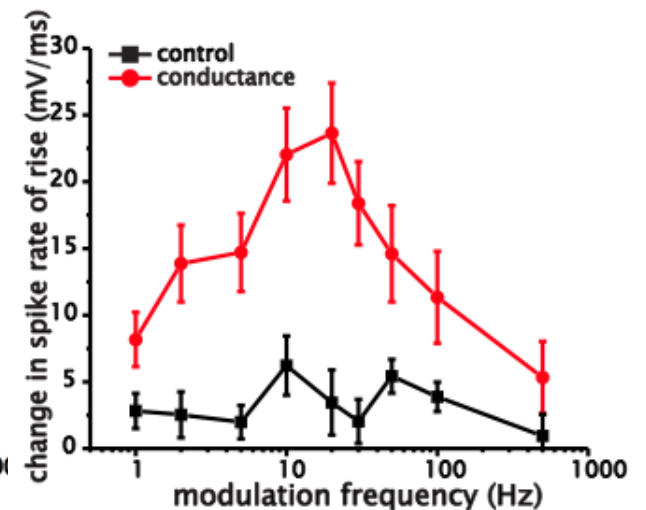
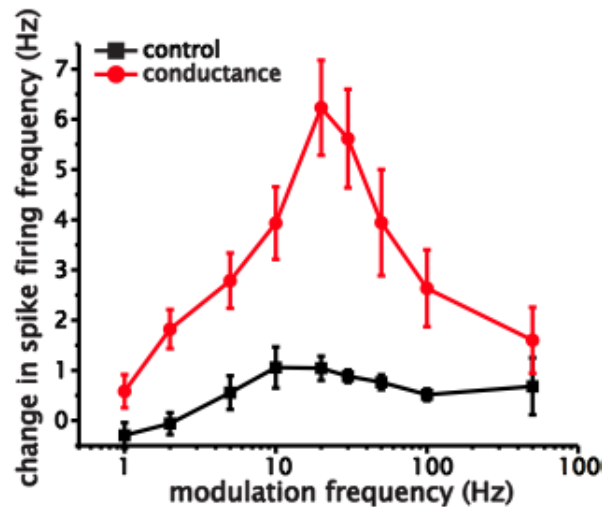
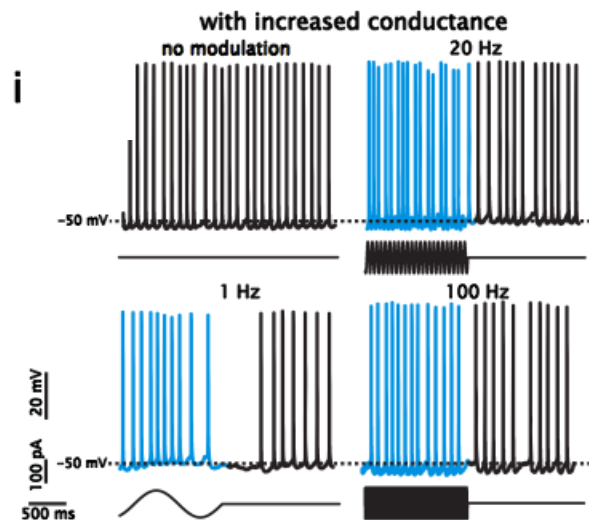
Chrobak and Buzsáki (1998) *J. Neurosci.* 18: 388-398

# Is coherent activity important for mnemonic function?

- Synchronous activity appears during learning
- Blocking synchrony blocks memory
- Epochs of correlated pre- and postsynaptic activity change the effective synaptic weight  
e.g. Keck et al. (2008) *J Neurosci* 28: 7359-7369
- Synaptic plasticity depends on timing  
e.g. Zhou et al. (2005) *PNAS USA* 102: 19121-19125
- Bouts of correlated activity affect postsynaptic gain  
e.g. Fernandez\*, Broicher\* et al. (2011) *J Neurosci* 31:3880-3993



# Periodic perturbations “rescue” inactivated $\text{Na}^+$ channels and restore gain in CA1 PCs

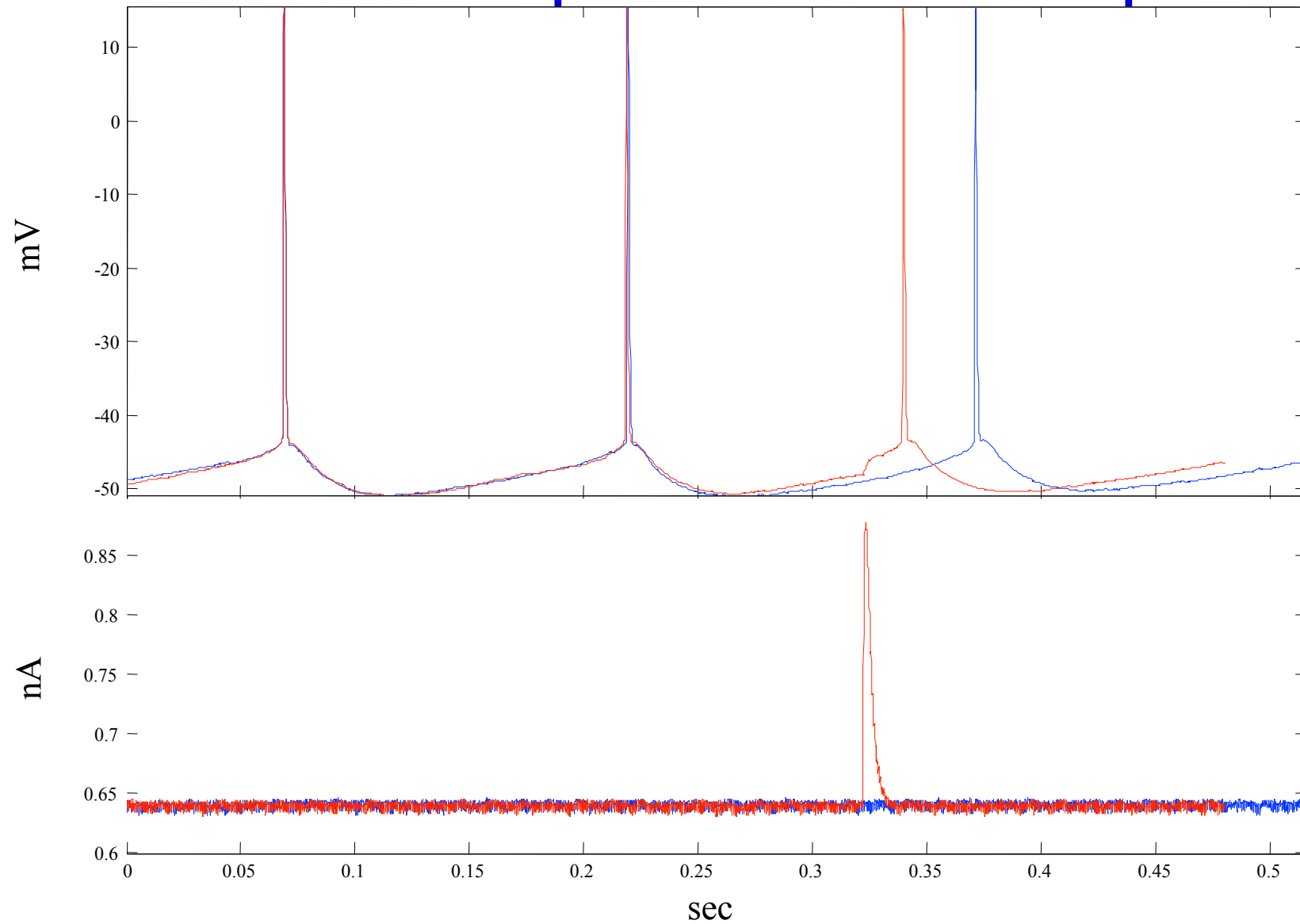


Fernandez\*, Broicher\* et al. (2011) *J Neurosci*

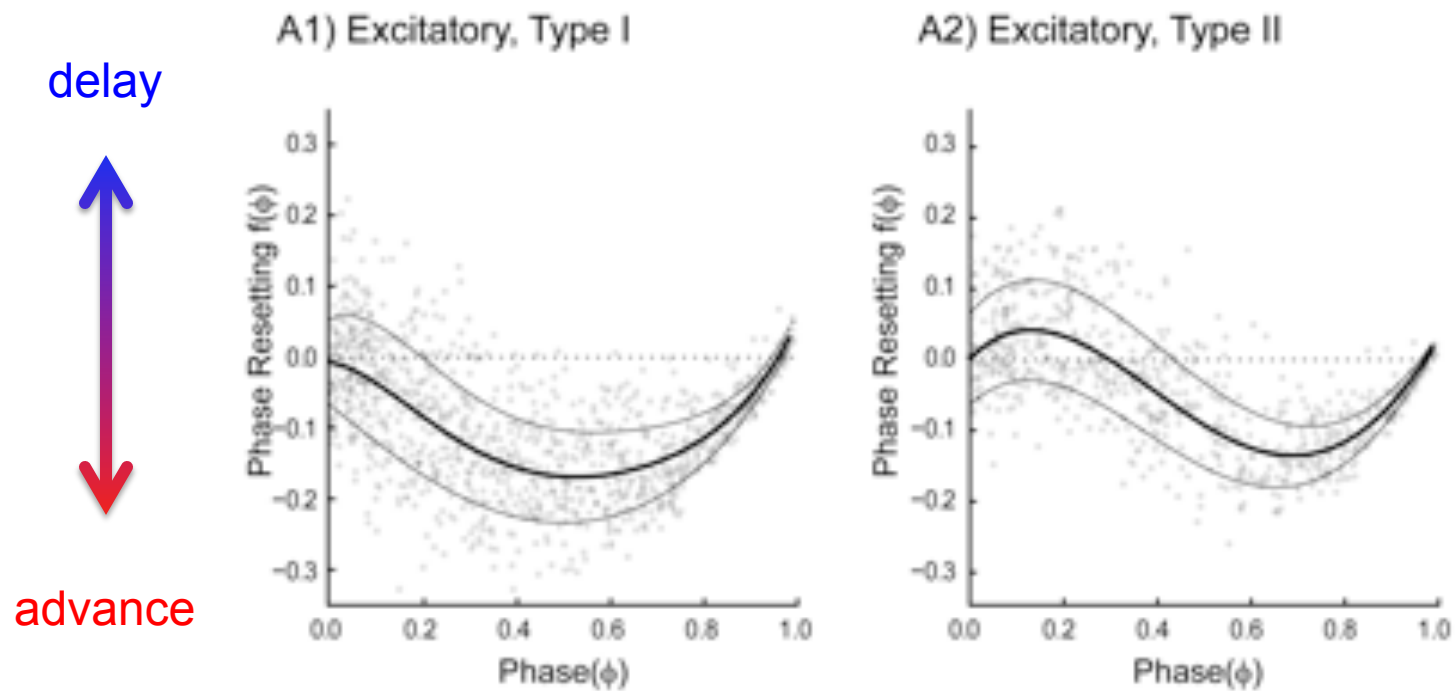
## Applications: Phase-response curves

- Cells act like oscillators
- Inputs perturb phase, with short-lived effects: *phase-response curve* describes this effect
- Helpful, but not necessary, if effects can be superimposed

# Phase response relationships

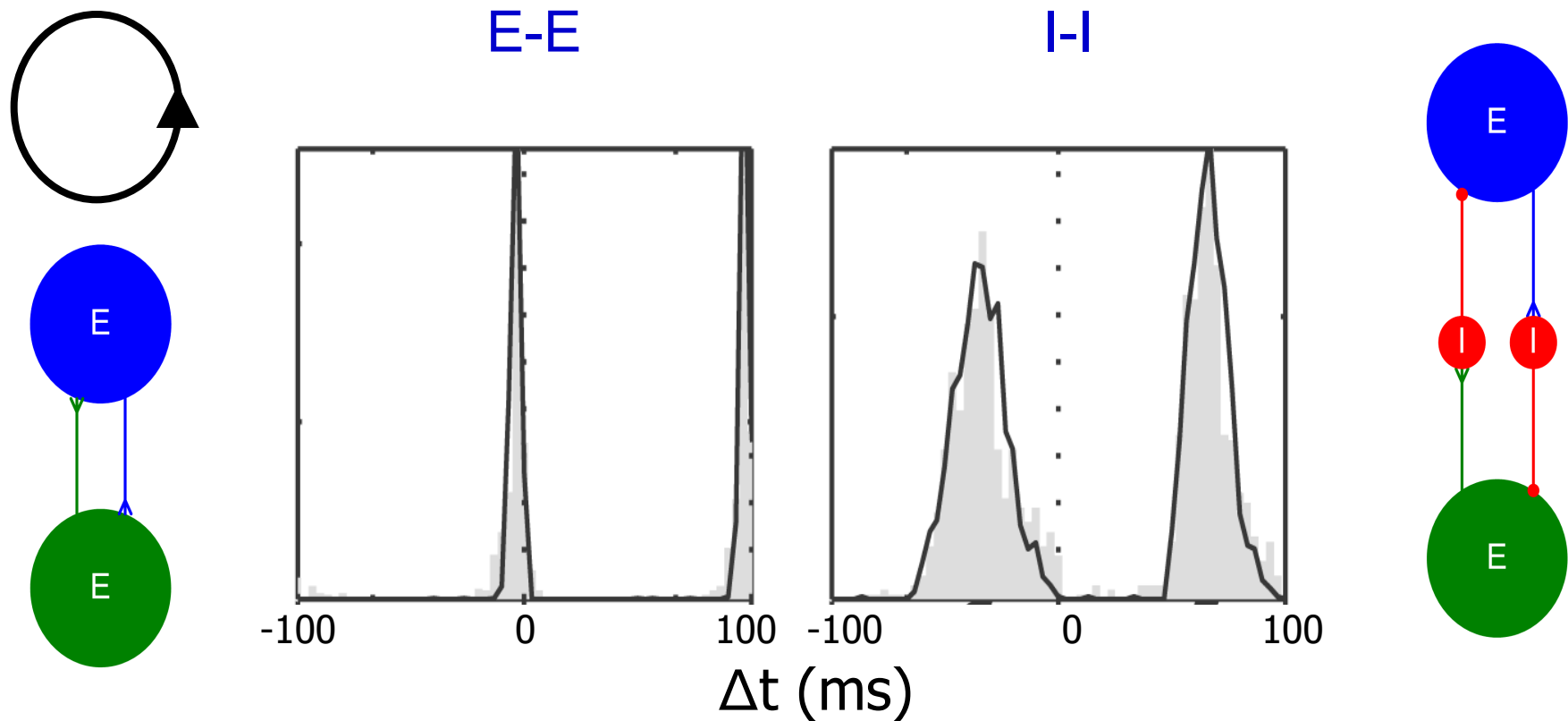


# Example PRCs: Entorhinal stellate cells



Data collected by Fernando Fernandez,  
in collaboration with Carmen Canavier

# SCs synchronize via mutual excitation, anti-synchronize via mutual inhibition

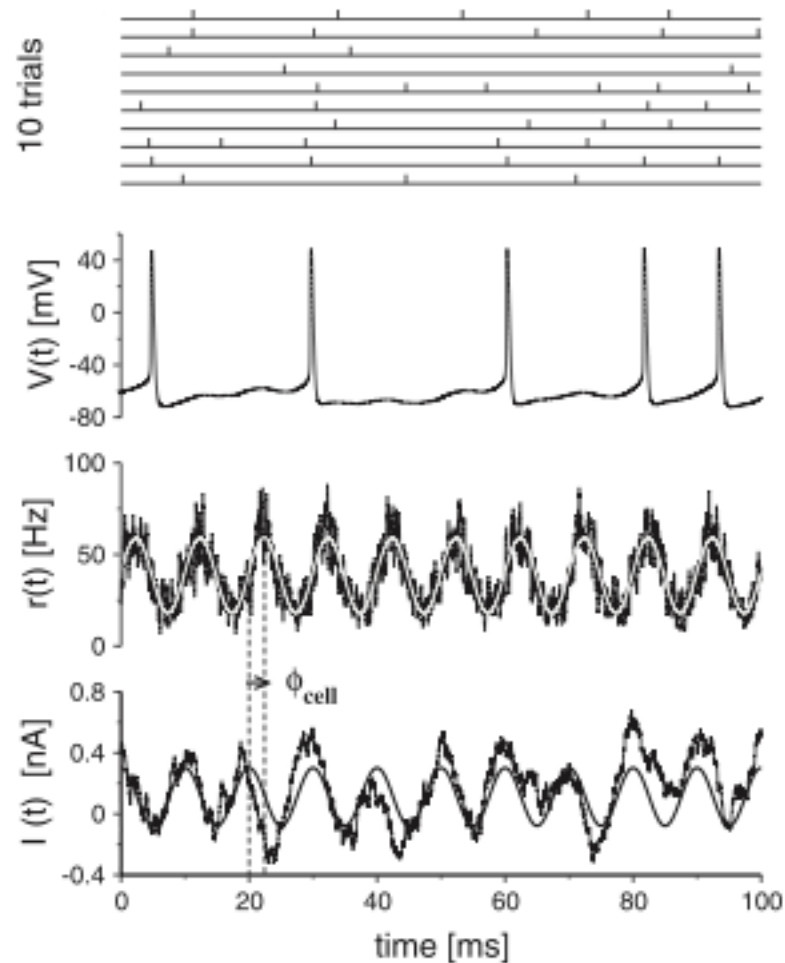


Netoff et al. (2005) *J Neurophysiol* 93: 1197-1208

Netoff et al. (2005) *J Comp Neurosci* 18: 287-295

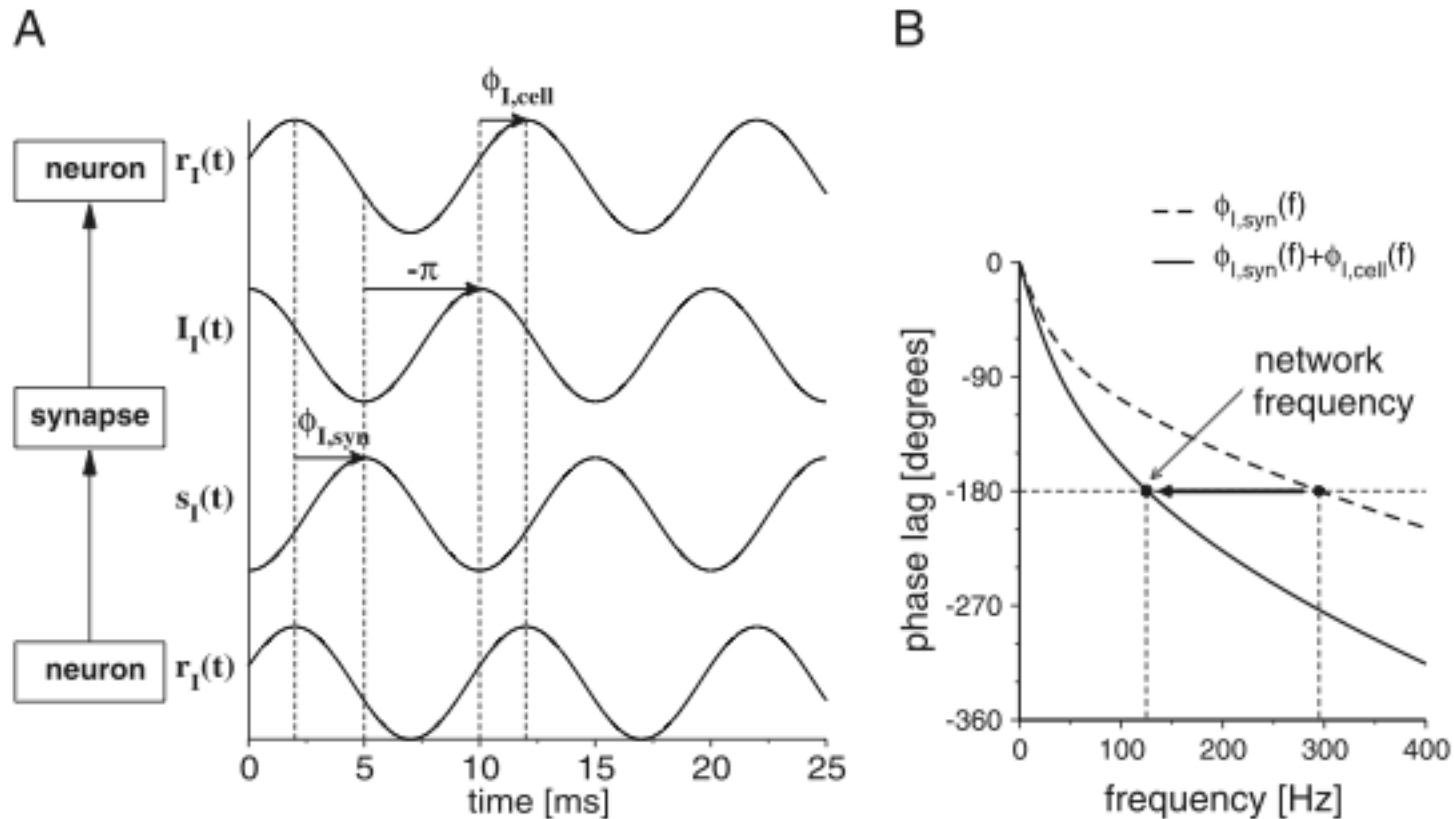
Pervouchine et al. (2006) *Neural Computation* 18:2617-2650

# Synchronization with irregular firing at low rates (1)



Geisler, Brunel, and Wang (2005) *J Neurophysiol* 94: 4344-4361

# Synchronization with irregular firing at low rates (2)





# Estimating $\phi_{I,\text{cell}}$ experimentally

